## Total Synthesis of 5(S),20- and 15(S),20-Dihydroxyeicosatetraenoic Acid and 5(S),6(R)-Epoxy-20-hydroxy- and 14(R),15(S)-Epoxy-20-hydroxyeicosatrienoic Acid

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Aldehyde (1), derived from dimethyl L-malate, was exploited in a stereocontrolled total synthesis of the  $\omega$ -hydroxylated eicosanoid metabolites 5(*S*),20- and 15(*S*),20-dihydroxyeicosatetraenoic acid as well as 5(*S*),6(*R*)-epoxy-20-hydroxy- and 14(*R*),15(*S*)-epoxy-20-hydroxyeicosatrienoic acid.

Cytochrome P-450 mediated  $\omega$ -hydroxylation is a major pathway in the secondary metabolism of eicosanoids<sup>1</sup> and can dramatically alter biological activity.<sup>2</sup> Recent examples include the oxidation of 12(S)-hydroxyeicosatetraenoic acid (HETE)<sup>3</sup> and several epoxygenase metabolites.<sup>4</sup> In vitro, cytochrome P-450 is also able to  $\omega$ -hydroxylate epoxyeicosatrienoic acids (EETs) in addition to other HETEs.<sup>†</sup> As part of our efforts<sup>5</sup> to evaluate the pharmacological consequences of  $\omega$ -hydroxylation and in anticipation of the isolation of new metabolites, a versatile and convergent enantiospecific approach to mono-oxygenase fatty acid metabolites was developed. The synthetic potential of this route is illustrated herein by total syntheses of 5(S),20- and 15(S),20-dihydroxyeicosatetraenoic acids as well as 5(S),6(R)-epoxy-20-hydroxyand 14(R),15(S)-epoxy-20-hydroxyeicosatrienoic acids.

Aldehyde (1)<sup>6</sup> was transformed into hydroxy-benzoate (2)‡ (61%) by homologation with 4-methoxycarbonylbutylidenetriphenylphosphorane under *cis*-olefination conditions, selective acetonide hydrolysis, and benzoylation of the primary alcohol (Scheme 1). Erythrospecific epoxidation<sup>7</sup> furnished (3) (77%) which was converted into aldehyde (4) (88%) by exposure to methanolic KHCO<sub>3</sub> followed by periodate cleavage of the resultant diol. Stirring with an ethereal suspension of silica gel smoothly isomerized (4) to the *trans*-enal (5) (96%).



Scheme 1. Reagents: i, Ph<sub>3</sub>PCH[CH<sub>2</sub>]<sub>3</sub>CO<sub>2</sub>Me, tetrahydrofuran (THF)-hexamethylphosphoric triamide (HMPT) (4:1),  $-78 \rightarrow 20^{\circ}$ C, 3 h; ii, 1 M HCl-MeOH (1:5), 4°C, 12 h; iii, PhCOCN, 10 mol% Et<sub>3</sub>N, CH<sub>2</sub>Cl<sub>2</sub>, 0°C, 0.5 h; iv, Bu'OOH, VO(acac)<sub>2</sub> (Hacac = MeCOCH<sub>2</sub>COMe), CH<sub>2</sub>Cl<sub>2</sub>, 0°C, 10 h; v, KHCO<sub>3</sub>, MeOH, 20 h; vi, NaIO<sub>4</sub>, MeOH-H<sub>2</sub>O (3:1), Na<sub>2</sub>HPO<sub>4</sub>, 0°C, 40 min; vii, SiO<sub>2</sub>, Et<sub>2</sub>O, 1 h.

<sup>&</sup>lt;sup>†</sup> The details of these experiments will be published elsewhere.

<sup>&</sup>lt;sup>‡</sup> Satisfactory n.m.r., u.v., and mass spectral data were obtained for all new compounds using chromatographically homogeneous samples.



Wittig coupling of (4) with 12-(t-butyldiphenylsilyloxy)dodeca-(Z,Z)-3,6-dien-1-ylidenetriphenylphosphorane [generated at -78 °C, tetrahydrofuran (THF), 45 min, NaN(SiMe<sub>3</sub>)<sub>2</sub>] in THF-toluene (1:4.6) at -100 to -15 °C over 4 h and fluoride mediated desilylation afforded methyl 5(S), 6(R)-epoxy-20-hydroxyeicosatrienoate (8) (59%).§ Treatment of (5) with the same phosphorane (-78 to -15 °C, 3 h) in THF-hexamethylphosphoric triamide (HMPT) (4:1) led to methyl 5(S), 20-dihydroxyeicosatetraenoate (9) (58%).

When the adduct between aldehyde (1) and 6-(tbutyldiphenylsilyloxy)hexylidenetriphenylphosphorane was carried through the sequence outlined in Scheme 1, aldehydes (6) and (7) were obtained in 44% and 42% overall yield, respectively. Condensation of (6) with 10-methoxycarbonyl-deca-(Z,Z)-3,6-dien-1-ylidenetriphenylphosphorane according to the conditions described above and desilylation resulted in methyl 14(R),15(S)-epoxy-20-hydroxyeicosatrienoate (10) (54%) whereas (7) gave methyl 15(S),20-dihydroxyeicosatetraenoate (11) (68%). Esters (8)—(11) were converted into the corresponding acids using LiOH in MeOH-H<sub>2</sub>O (3:1), acidification to pH 4.5, and extractive isolation.

Results from current investigations into the occurrence, enzymatic origin, and pharmacology of the above metabolites will be reported later.

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<sup>§ (8): &</sup>lt;sup>1</sup>H n.m.r. (90 MHz, CDCl<sub>3</sub>)  $\delta$  1.20–2.48 (16H, m), 2.66–3.02 (6H, m), 3.58 (2H, t, *J* 7 Hz), 3.62 (3H, s), 5.20–5.56 (6H, m);  $[\alpha]_D^{23}$  -3.77° (*c* 1.28, acetone). (9): <sup>1</sup>H n.m.r. (90 MHz, CDCl<sub>3</sub>)  $\delta$  1.16–1.72 (10H, m), 1.90–2.44 (4H, m), 2.66–3.02 (4H, m), 3.57 (2H, t, *J* 7 Hz), 3.62 (3H, s), 4.00–4.24 (1H, m), 5.14–6.05 (7H, m), 6.46 (1H, dd, *J* 11 and 15 Hz);  $[\alpha]_D^{23}$  10.2° (*c* 1.53, acetone). (10):  $[\alpha]_D^{23}$  4.66° (*c* 1.01, acetone). (11):  $[\alpha]_D^{23}$  12.0° (*c* 1.76, acetone). The <sup>1</sup>H n.m.r. spectra of (10) and (11) are virtually identical to those of (8) and (9), respectively.